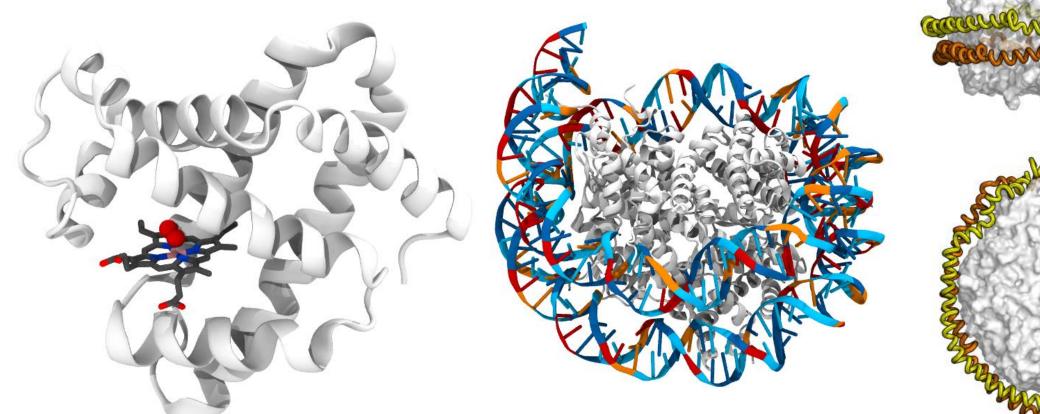
## Molecular Biophysics

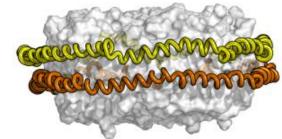
Protein assembly, function and malfunction

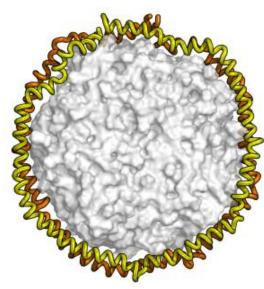
#### **Summary of last lessons**

- Proteins are amino acid polymers that fold in a 3D structure
- amino acid sequence + environment = conformational space
- The structure determines the function: proteins interact with specific binding partners
- Means to study protein structure: X-ray crystallography, Nuclear Magnetic Resonance (NMR), Electron Microscopy, Modelling
- Our view of the proteome is biased by the techniques we use to observe it

## Proteins interact with specific binding partners







Myoglobin (PDB: 1MBO)

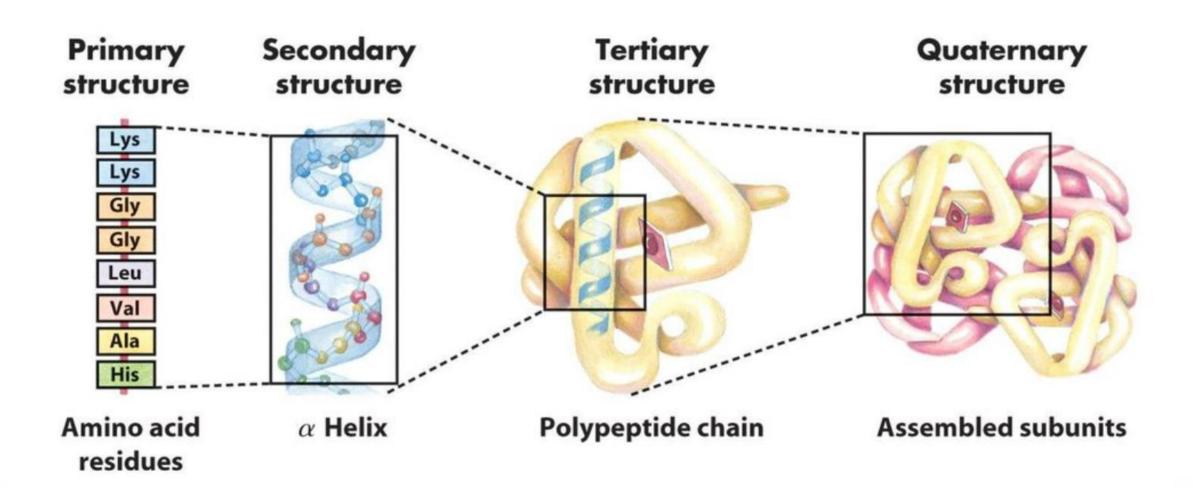
J.C. Kendrew et al., A three-dimensional Model of the Myoglobin Molecule obtained by X-Ray Analysis, Nature, 1958 Nucleosome (PDB: 5CPI)

K. Luger et al., *Crystal Structure of the nucleosome core particle at 2.8 A resolution,* Nature, 1997

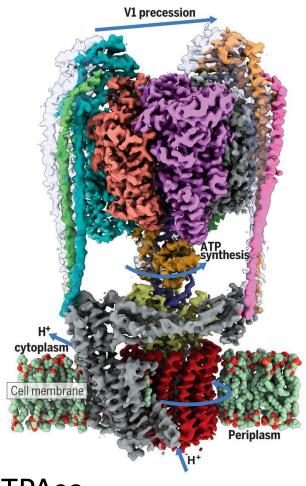
Lipoprotein (PDB: 1AV1)

D.W. Bohrani et al., *Crystal structure of truncated human apolipoprotein A-I suggests a lipid-bound conformation*, PNAS, 1997

#### Protein quaternary structure: assembly

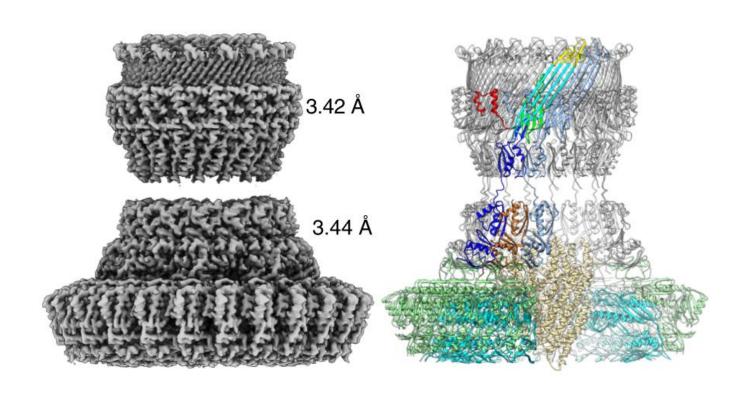


#### Protein quaternary structure: assembly



**ATPAse** 

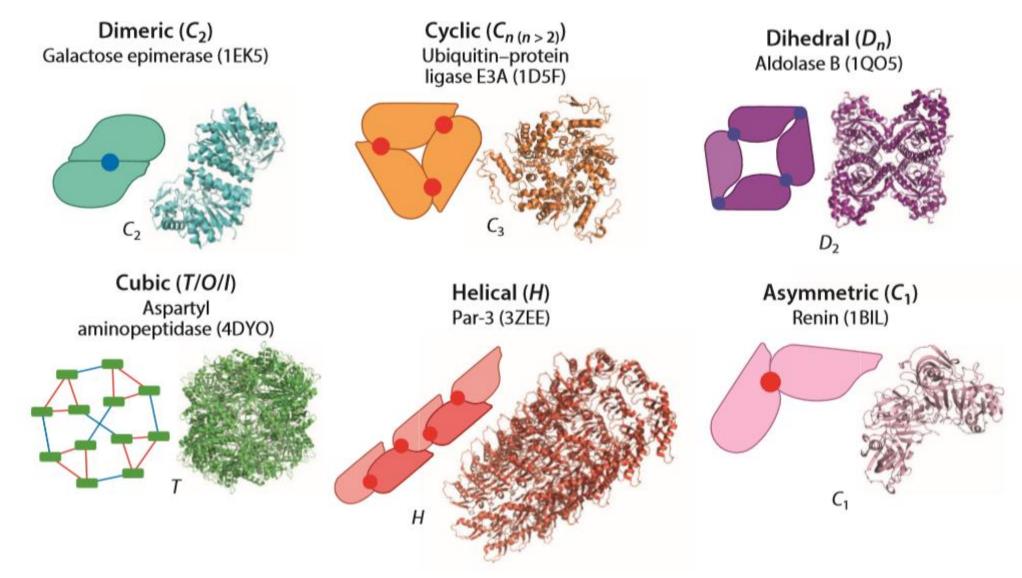
L. Zhou and L. A. Sazanov, *Structure and* conformational plasticity of the intact Thermus thermophilus V/A-type ATPase, Science, 2019



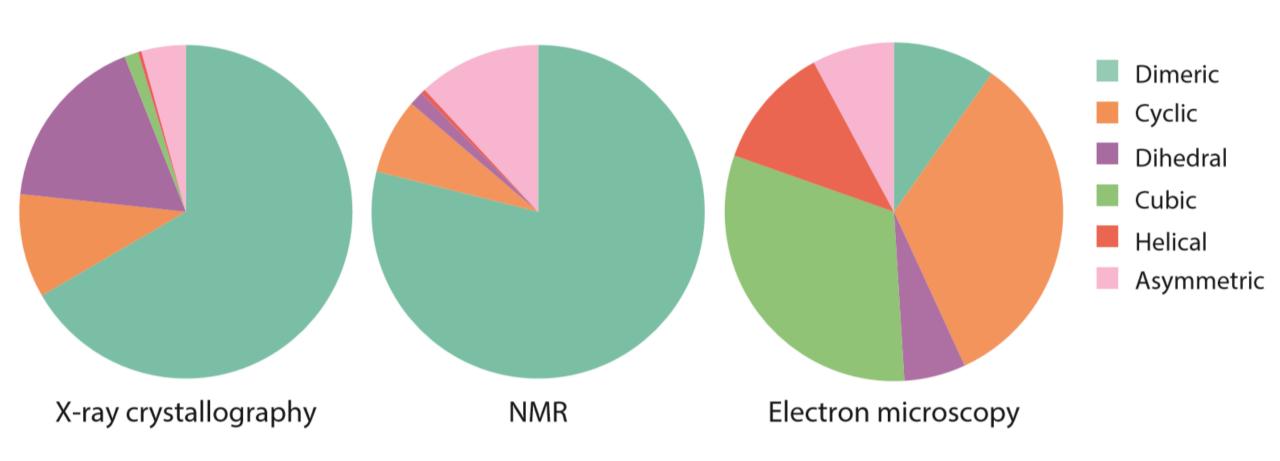
Injectisome (evolutionarily related to the flagellum)

J.Hu et al., *Cryo-EM analysis of the T3S injectisome* reveals the structure of the needle and open secretin, Nature comms, 2018

### Assembly stoichiometry and symmetry



#### Assembly stoichiometry and symmetry



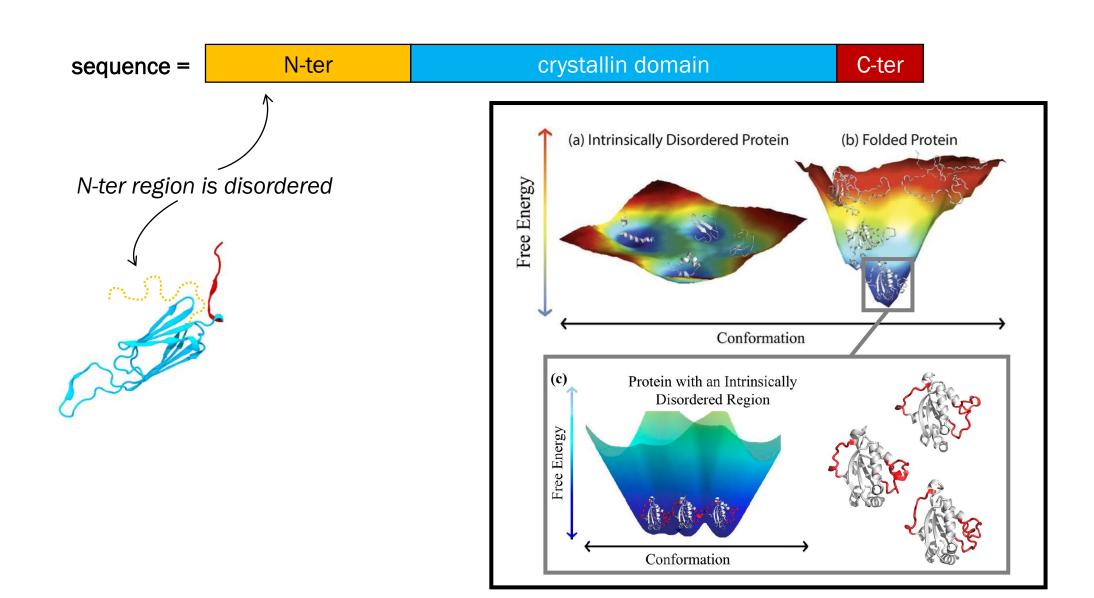
# Small heat-shock protein (HSP): countering protein unfolding

<u>Denaturation</u>: protein unfolding caused by its exposition to non-native conditions, e.g.:

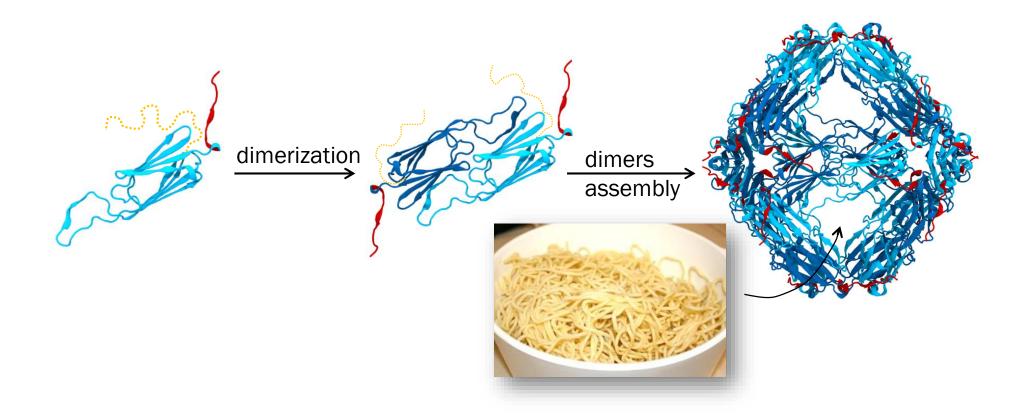
- heat
- pH
- reducing/oxydising agents

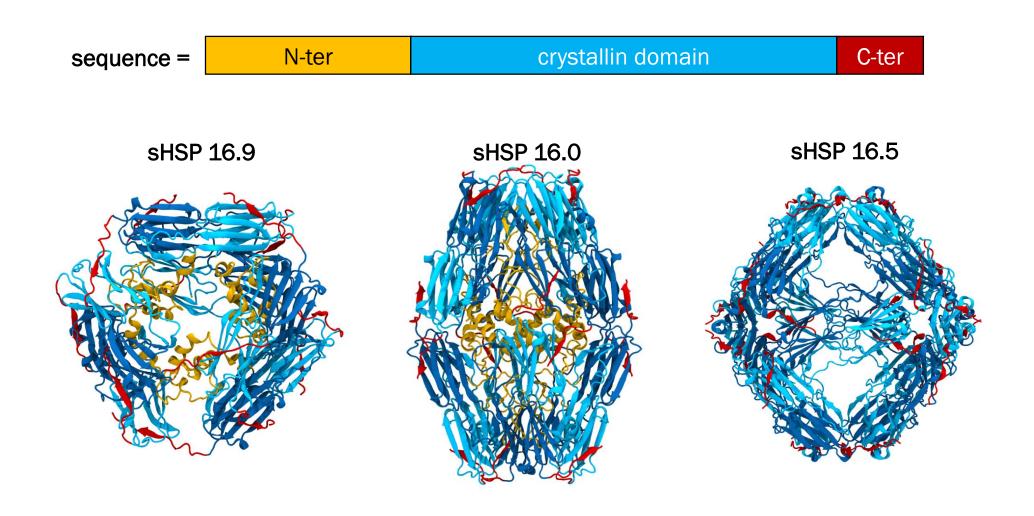


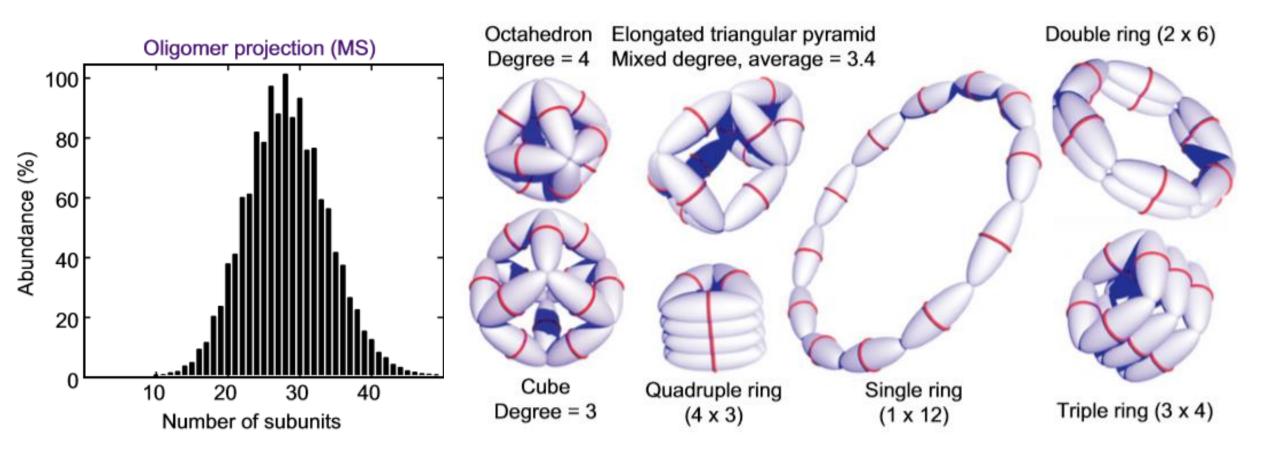
Protein denaturation is usually harmful for an organism



sequence = N-ter crystallin domain C-ter



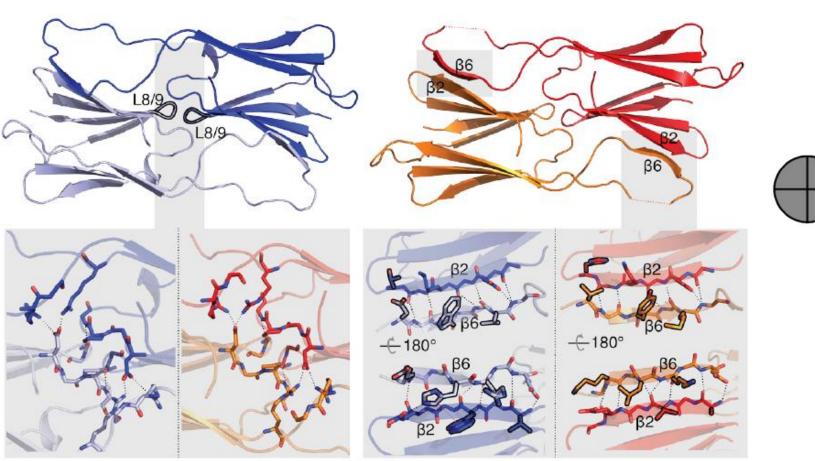


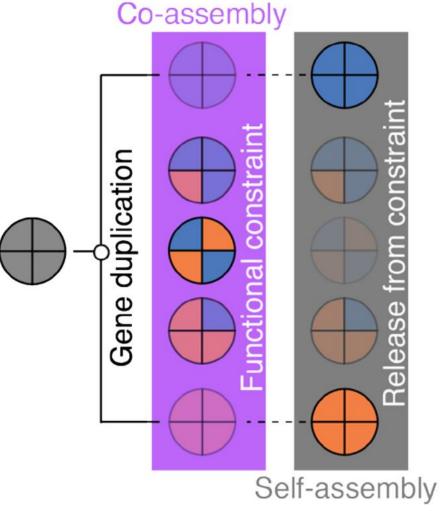


A.J. Baldwin at al., The Polydispersity of αB-Crystallin Is Rationalized by an Interconverting Polyhedral Architecture, Structure, 2011 I. Santhanagopalan et al, It takes a dimer to tango: Oligomeric small heat shock proteins dissociate to capture substrate, JBC, 2018

#### HSP: similars don't co-assemble

 High sequence homology, near-identical dimers interfaces (1.2 Å backbone RMSD)

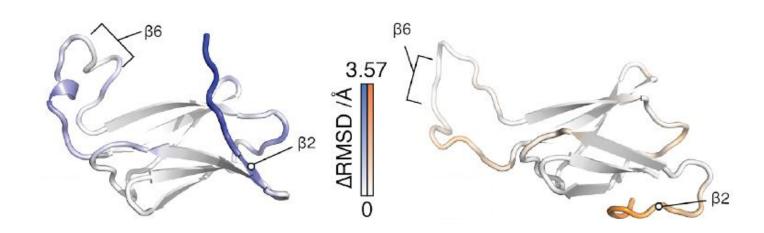


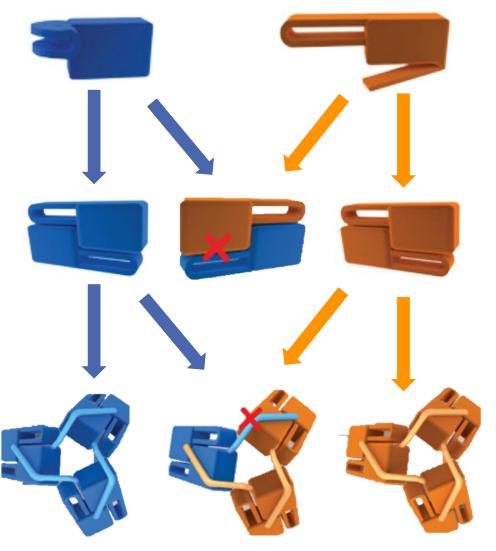


G.K. Hochberg et al. Structural principles that enable oligomeric small heat-shock protein paralogs to evolve distinct functions. Science, 2018

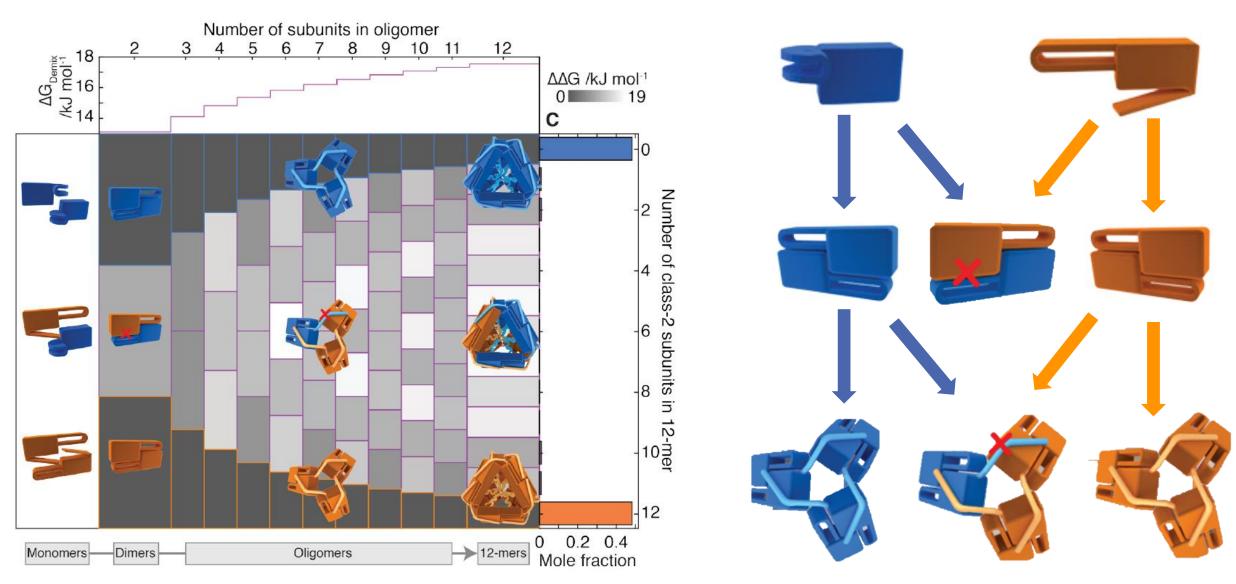
## HSP: selectivity is mediated by dynamics

- High sequence homology, near-identical dimers interfaces (1.2 Å backbone RMSD)
- Monomeric subunits feature different disordered regions

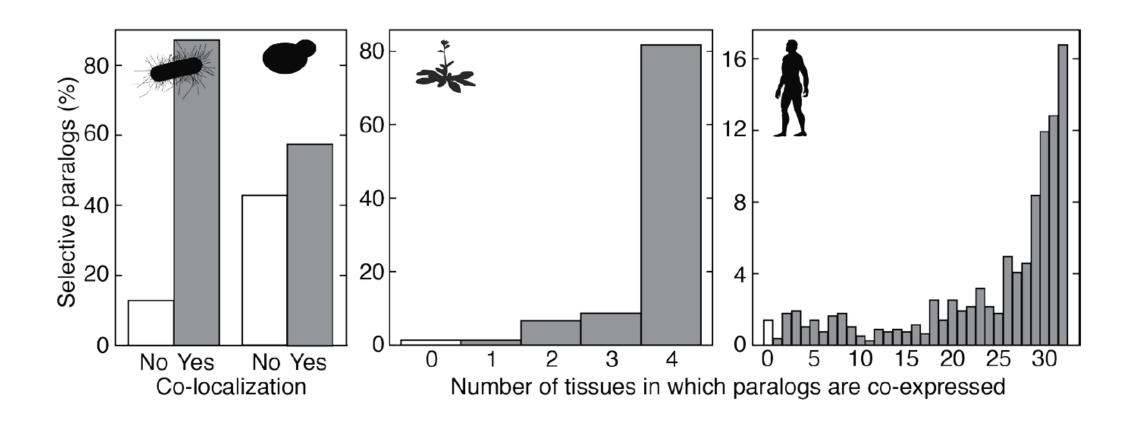




## HSP: selectivity overcomes entropy



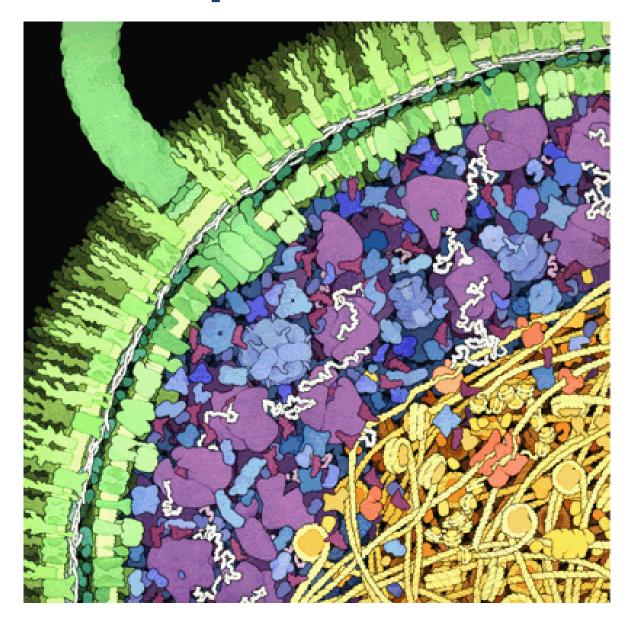
## [Extra] Most paralogs co-localise but don't co-assemble



#### The intracellular space

A crowded environment, Brownian motion: proteins bump into each other all the time!

Most contacts are non-specific and short-lived



reported proteins per cell	cell volume (μm³)	proteins per volume (10 <sup>6</sup> /μm <sup>3</sup>
M. pneumonia		
0.05×10 <sup>6</sup>	0.015	3
L. interrogans		
1.0-1.2×10 <sup>6</sup> *	0.22	5
E. coli		
2.36×10 <sup>6</sup>	0.86	2.7
B. subtilis		
2.3×10 <sup>6</sup> *	1.13	2.0
1.3×10 <sup>6</sup> *	0.62	2.1
1.8×10 <sup>6</sup> *	0.85	2.1
S. aureus		
0.35×10 <sup>6</sup> *	0.33	1.1
0.27×10 <sup>6</sup> *	0.23	1.2
0.26×10 <sup>6</sup> *	0.23	1.1
budding yeast (haploid)		
50×10 <sup>6</sup>	≈30-40	1-2
47×10 <sup>6</sup> *	≈30-40	1-2
53×10 <sup>6</sup>	≈30-40	1-2
fission yeast		
60.3×10 <sup>6</sup>	≈100	0.6
M. musculus (NIH3T3 cells)		
3×10 <sup>9</sup> *	≈2000	1.5
H. Sapiens (U2OS)		
0.95-1.7×10 <sup>9</sup> *	≈4000	0.2-0.4
H. sapiens (HeLa)		
2.0×10 <sup>9</sup> *	≈2000	1

### **Protein density**

Estimate of proteins per unit of volume  $N_V$ 

$$N_V = \frac{N}{V} = \frac{m_{all}}{m_{prot}} N_A \frac{1}{V} = \frac{N_A}{n_{aa} m_{aa}} \frac{m_{all}}{V}$$

 $m_{all}$ : total protein mass in cell (g)  $n_{aa}$ : average #aminoacids per  $m_{prot}$ : avg. protein molar mass (g/mole) protein  $\approx$  **300**  $m_{aa}$ : average mass of one  $m_{aa}$ : average mass of one aminoacid  $\approx$  **100 Da** 

• Typical protein mass inside cells  $(m_{all}/V)$ : 200-300 g/L. ~20-30% of total volume!

molecule	measured context	diffusion coefficient (µm²/s)
H <sub>2</sub> O	water	2000
H <sub>2</sub> O	nucleus of chicken erythrocyte	200
$H^+$ (from $H_3O^+$ to $H_2O$ )	water	7000
O <sub>2</sub>	water	2000
CO <sub>2</sub>	water	2000
tRNA (≈20 kDa)	water	100
protein (≈30 kDa GFP)	water	100
protein (≈30 kDa GFP )	eukaryotic cell (CHO) cytoplasm	30
protein (≈30 kDa GFP )	rat liver mitochondria	30
protein (NLS-EGFP)	cytoplasm of <i>D. melanogaster</i> embryo	20
protein (≈30 kDa )	E. coli cytoplasm	7-8
protein (≈40 kDa )	E. coli cytoplasm	2-4
protein (≈70-250 kDa )	E. coli cytoplasm	0.4-2
protein (≈140 kDa Tar-YFP)	E. coli membrane	0.2
protein (≈70 kDa LacY-YFP)	E. coli membrane	0.03
fluorescent dye (carboxy-fluorescein)	A. thaliana cell wall	30
fluorescent dye (carboxy-fluorescein)	A. thaliana mature root epidermis	3
transcription factor (Lacl)	movement along DNA (1D, in vitro)	$0.04$ $(4\times10^5 \text{ bp}^2\text{s}^{-1})$
morphogen (bicoid-GFP)	cytoplasm of <i>D. melanogaster</i> embryo	7
morphogen (wingless)	wing imaginal disk of D. melanogaster	0.05
mRNA	HeLa nucleus	0.03-0.10
mRNA	various localizations and sizes	0.005-1
ribosome	E. coli	0.04

#### **Protein diffusion**

#### Stokes-Einstein relation:

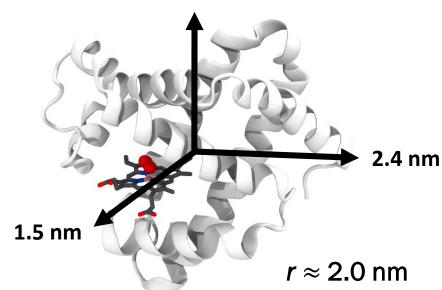
$$D = \frac{k_B T}{6\pi \eta r}$$

r: radius of a protein (nm)

T: temperature (K)

 $\eta$ : viscosity (N s/m<sup>2</sup>)

#### 2.1 nm



#### In the cytoplasm:

$$T \approx 300 \text{ K}$$
  
 $\eta \approx 10^{-2} \text{ N s/m}^2$ 

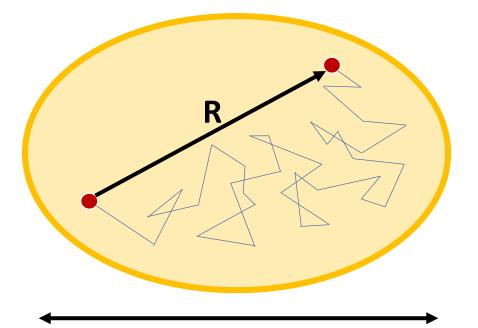
$$D \approx 10^{\mu m^2}/_{s}$$

book.bionumbers.org/what-are-the-time-scales-for-diffusion-in-cells

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ribosome	E. coli	0.04

#### **Protein diffusion**

$$D \approx 10^{\mu m^2}/_{S}$$



*E. coli* R  $\approx$  1  $\mu m$ 

$$t = \frac{R^2}{6D} \approx 17 \, ms$$

book.bionumbers.org/what-are-the-time-scales-for-diffusion-in-cells

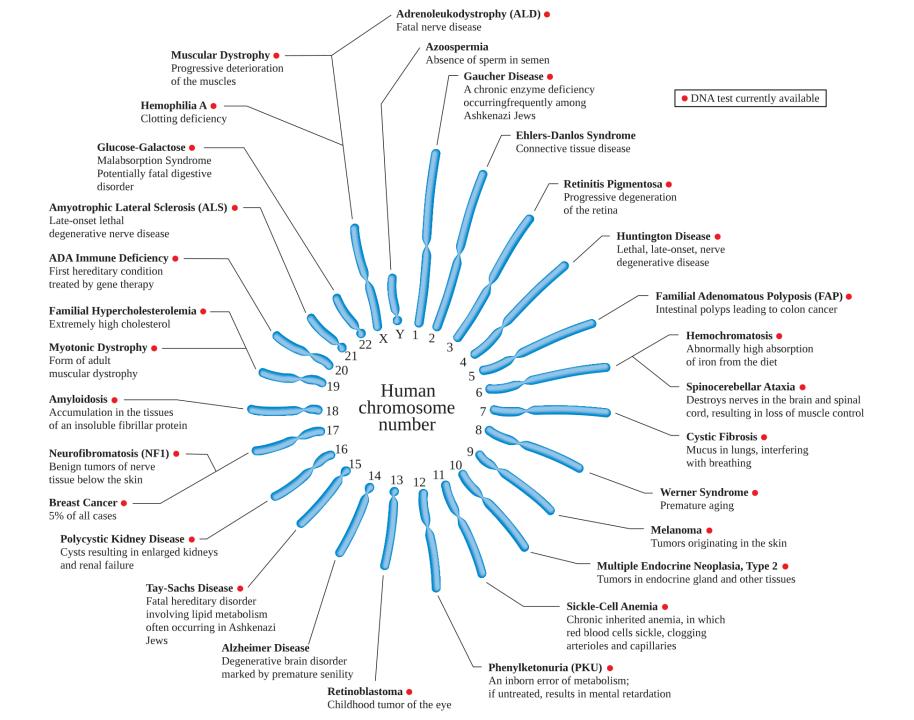
#### Protein function and malfunction

#### Effect of mutations

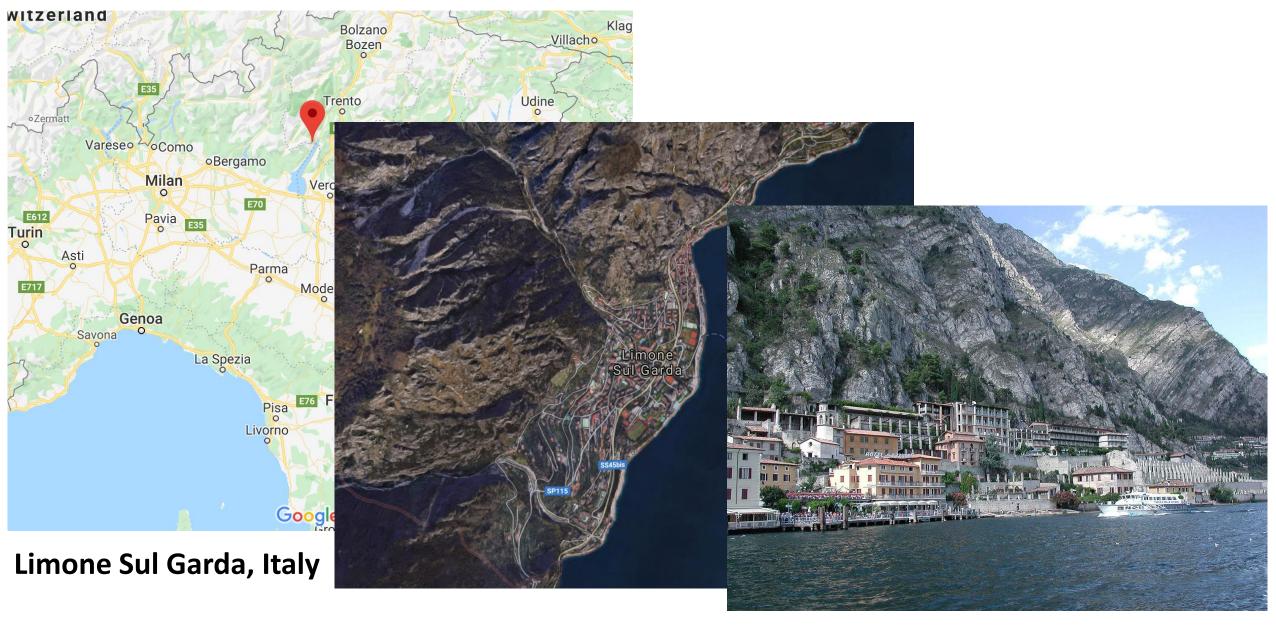
- Protein cannot fold
- Proteins misfolds
- Protein folds, but has different dynamics

#### Consequences of mutations

- Protein cannot bind to anything
- Protein interacts with one or more different binding partners



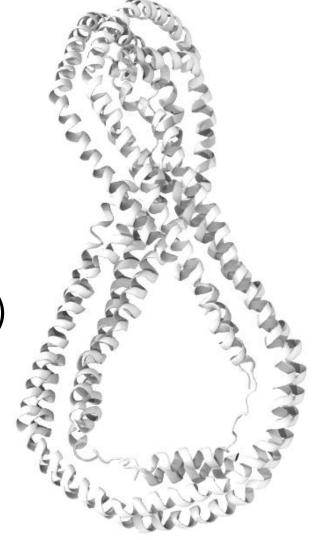
#### Sometimes mutations are beneficial



#### Sometimes mutations are beneficial

#### apolipoprotein A-1 Milano:

- carries cholesterol from tissues to the liver
- Phenotype:
  - reduction in HDL levels and an increase in triglyceride levels
  - positive effects against atherosclerosis
- mutation R173C developed by Giovanni Pomarelli (1780)
- gene therapy works well on animal models
- research as medical treatment stalled (Esperion, Pfizer, SemBioSys, ...)



blogs.sciencemag.org/pipeline/archives/2016/11/16/the-long-saga-of-apo-a1-milano

#### Take home messages

Proteins often assemble into complexes to carry out their task

The intracellular space is crowded

Mutation may alter protein capacity of interacting with its dedicated partner